

**2006-027: DRAFT ANNEX to ISPM 27– Sorghum halepense**

| Comm . no. | Para . no. | Comment type | Comment | Explanation | Country | SC response |
|------------|------------|--------------|---|-------------|---|---|
| 1. | G | Editorial | <u>It is recommended that this protocol paragraphs are numbered for clarity and document management.</u> | Clarify | Costa Rica | Noted. The IPPC Secretariat will adjust the formatting once the DP is adopted. |
| 2. | G | Substantive | I support the document as it is and I have no comments | | Georgia, Singapore, New Zealand, Nepal, Mexico, Congo, South Africa, Barbados, Bahrain, Guyana, Belize, Ghana, Burundi | Noted |
| 3. | G | Technical | <p><u>1. We would like to request the TPDP to include in this diagnostic protocol the uncertainty level of each method described in section 4, if available, in order to know their level of analytical confidence. We would also like the TPDP to consider the possibility to include a comparative table containing all methods with their uncertainty levels.</u></p> <p><u>2. We suggest the TPDP to reflect in this DP that morphological identification of seeds should be complemented by the morphological identification of plants as diagnostic confirmation test.</u></p> <p><u>3. QBOL is a consortium of 20 partners (universities, research institutes and phytosanitary organizations) from all over the world working together and sharing their research expertise in the field of DNA</u></p> | See comment | Peru | <p>1. Incorporate. A comparative table on each of identification method was included</p> <p>2. Incorporated. Revision done to paragraph 29</p> <p>3. Considered but not incorporated. However, we welcome to make more comments and suggestions</p> |



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| | | | <u>barcoding of Arthropods, Bacteria, Fungi, Nematodes, Phytoplasmas and Viruses. Thereby, we would like to request the TPDP to evaluate the relevance to include this method in protocols.</u> ✖ | | | |
| 4. | G | Technical | <p><u>1. We would like to request the TPDP to include in this diagnostic protocol the uncertainty level of each method described in section 4, if available, in order to know their level of analytical confidence. We would also like the TPDP to consider the possibility to include a comparative table containing all methods with their uncertainty levels.</u></p> <p><u>2. We suggest the TPDP to reflect in this DP that morphological identification of seeds should be complemented by the morphological identification of plants as diagnostic confirmation test.</u> ✖</p> | See comment | Argentina | 1. Incorporated. See comment 3.2. Incorporated. See comment 3. |
| 5. | G | Technical | <p><u>1. We would like to request the TPDP to include in this diagnostic protocol the uncertainty level of each method described in section 4, if available, in order to know their level of analytical confidence. We would also like the TPDP to consider the possibility to include a comparative table containing all methods with their uncertainty levels.</u></p> <p><u>2. We suggest the TPDP to reflect in this DP that morphological identification of seeds should be complemented by the morphological identification of plants as diagnostic confirmation test.</u></p> <p>✖</p> | See comment | COSAVE | <p>1. Incorporated. See comment 3.</p> <p>2. Incorporated. See comment 3.</p> <p>3. Considered but not incorporated. However, we welcome to make more comments and suggestions (see comment 3)</p> |
| 6. | G | Technical | <u>1. We would like to request the TPDP to include in this diagnostic protocol the uncertainty level of each method described in section 4, if available, in order to know their level of analytical confidence. We would also like the TPDP to consider the possibility to include a comparative table containing all methods with their uncertainty levels.</u> | See comment | Brazil | <p>1. Incorporated. See comment 3.</p> <p>2. Incorporated. See comment 3.</p> |



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| | | | <p><u>2. We suggest the TPDP to reflect in this DP that morphological identification of seeds should be complemented by the morphological identification of plants as diagnostic confirmation test.</u></p> <p><u>3. QBOL is a consortium of 20 partners (universities, research institutes and phytosanitary organizations) from all over the world working together and sharing their research expertise in field of DNA barcoding of Arthropods, Bacteria, Fungi, Nematodes, Phytoplasmas and Virus. Thereby, we would like to request the TPDP to evaluate the relevance to include this method in this protocol.</u></p> | | | 3. Considered but not incorporated. However, we welcome to make more comments and suggestions (see comment 3) |
| 5. | G | Technical | <p><u>1. We would like to request the TPDP to include in this diagnostic protocol the uncertainty level of each method described in section 4, if available, in order to know their level of analytical confidence. We would also like the TPDP to consider the possibility to include a comparative table containing all methods with their uncertainty levels.</u></p> <p><u>2. We suggest the TPDP to reflect in this DP that morphological identification of seeds should be complemented by the morphological identification of plants as diagnostic confirmation test.</u></p> | See comment | Uruguay, Chile, Paraguay | <p>1. Incorporated. See comment 3.</p> <p>2. Incorporated. See comment 3.</p> |
| 8. | 8 | Substantive | 1. Pest Information | Reference to essential reviews on this pest are missing, e.g. Warwick et al. (1993) Canadian Journal of Plant Science 63: 997-1014. | EPPPO | Incorporated. Referred to it, improved the description |
| 9. | 8 | Substantive | 1. Pest Information <u>Include information on seed description.</u> | It would be helpful to include a brief description of the seed, including seed size under pest information. This will provide context for 'Section 3.2. Sieve detection'. | Australia | Incorporated. Referred to it, improved the description |
| 10. | 8 | Substantive | 1. Pest Information | References to essential reviews on this pest are missing, e.g. Warwick&Black (1983) Canadian Journal of Plant Science 63: 997- | European Union | Incorporated. |



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| | | | | 1014; Follak&Essl (2012) Weed Research 53(1):53-60. | | |
| 11. | 9 | Editorial | <p><i>Sorghum halepense</i> (Johnsongrass) is a perennial grass with a ribbed leaf sheath, conspicuous midrib, large, purplish panicles, and far-reaching rhizomes (Figures 1 and 2[no space between]). It originated from the hybridization of <i>Sorghum arundinaceum</i> and <i>Sorghum propinquum</i> through chromosome doubling (chromosomes: $2n = 4x = 40$) (Ng'uni et al., 2010).</p> <p><i>S. halepense</i> which is native to the Mediterranean area (Meredith, 1955) and was introduced to India in the late 1960s (Bor, 1960). It has become widespread, and is distributed from latitude 55° north to 45° south. It is best adapted to warm, humid areas with summer rainfall, areas with a high water table, and irrigated fields in subtropical zones. <i>S. halepense</i> is one of the most malignant weeds worldwide, impacting more than 30 cereal, vegetable and fruit crops (Holm <i>et al.</i>, 1977). It also threatens biodiversity in invaded habitats in no fewer than 50 countries in temperate and tropical areas throughout the world, including countries in which it is a native species (Holm <i>et al.</i>, 1977).</p> | grammatical correction | Kenya | Incorporated. |
| 12. | 9 | Substantive | <p><i>Sorghum halepense</i> (Johnsongrass) is a perennial grass with a ribbed leaf sheath, conspicuous midrib, large, purplish panicles, and far-reaching rhizomes (Figures 1 and 2-). Its <u>origin is uncertain, some authors suggest that it</u> originated from the hybridization of <i>Sorghum arundinaceum</i> and <i>Sorghum propinquum</i> through chromosome doubling (chromosomes: $2n = 4x = 40$) (Ng'uni et al., 2010).</p> <p><i>S. halepense</i> which is native to the Mediterranean area (Meredith, 1955) and was introduced to India in the late 1960s (Bor, 1960). It has become widespread, and is distributed from latitude 55° north to 45° south. It is best adapted to warm, humid</p> | the origin of <i>S. halepense</i> is not as clear as suggested here. Another possibility supported by the cited study and Morden et al. 1990 is that one of the parent of <i>Sorghum halepense</i> is <i>Sorghum bicolor</i> . It could be useful to give precisions (citing references)-: which habitats are concerned and where does this impact occur? | EPPO | <p>1. Modified.</p> <p>2. Modified. The mention to “in invaded habitats” has been removed from the text. However, the reference to Holm et al., 1997 remains.</p> |



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| | | | <p>areas with summer rainfall, areas with a high water table, and irrigated fields in subtropical zones. <i>S. halepense</i> is one of the most malignant weeds worldwide, impacting more than 30 cereal, vegetable and fruit crops (Holm <i>et al.</i>, 1977). It also threatens biodiversity in invaded habitats (which ones and how) in no fewer than 50 countries in temperate and tropical areas throughout the world, including countries in which it is a native species (Holm <i>et al.</i>, 1977).</p> | | | |
| 13. | 9 | Substantive | <p><i>Sorghum halepense</i> (Johnsongrass) is a perennial grass with a ribbed leaf sheath, conspicuous midrib, large, purplish panicles, and far-reaching rhizomes (Figures 1 and 2). Its origin is uncertain, some authors suggest that it originated from the hybridization of <i>Sorghum arundinaceum</i> and <i>Sorghum propinquum</i> through chromosome doubling (chromosomes: 2n = 4x = 40) (Ng'uni <i>et al.</i>, 2010).</p> <p><i>S. halepense</i> which is native to the Mediterranean area (Meredith, 1955) and was introduced to India in the late 1960s (Bor, 1960). It has become widespread, and is distributed from latitude 55° north to 45° south. It is best adapted to warm, humid areas with summer rainfall, areas with a high water table, and irrigated fields in subtropical zones. <i>S. halepense</i> is one of the most malignant weeds worldwide, impacting more than 30 cereal, vegetable and fruit crops (Holm <i>et al.</i>, 1977). It also threatens biodiversity in invaded habitats (which ones and how) in no fewer than 50 countries in temperate and tropical areas throughout the world, including countries in which it is a native species (Holm <i>et al.</i>, 1977).</p> | <p>The origin of <i>S. halepense</i> is not as clear as suggested here. Another possibility supported by the cited study and Morden <i>et al.</i> 1990 is that one of the parents of <i>Sorghum halepense</i> is <i>Sorghum bicolor</i>. It could be useful to give precisions (citing references) : which habitats are concerned and where does this impact occur?</p> | European Union | <p>Modified.</p> <p>See response above (comment 12)</p> |
| 14. | 9 | Technical | <p><i>Sorghum halepense</i> (Johnsongrass) is a perennial grass with a ribbed leaf sheath, conspicuous midrib, large, purplish panicles, and far-reaching rhizomes (Figures 1 and 2). It originated from the hybridization of <i>Sorghum arundinaceum</i> and</p> | <p>it is a bit strange to focus on India here since the species has become established in all the warm regions of the world as explained in the next sentence. Either</p> | EPPPO | <p>Incorporated (Modified).</p> <p>Text adjusted to "other regions then"</p> |



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| | | | <p><i>Sorghum propinquum</i> through chromosome doubling (chromosomes: $2n = 4x = 40$) (Ng'uni et al., 2010). <i>S. halepense</i> which is native to the Mediterranean area (Meredith, 1955) and was introduced to India in the late 1960s (Bor, 1960). It has become widespread, and is distributed from latitude 55° north to 45° south. It is best adapted to warm, humid areas with summer rainfall, areas with a high water table, and irrigated fields in subtropical zones. <i>S. halepense</i> is one of the most malignant weeds worldwide, impacting more than 30 cereal, vegetable and fruit crops (Holm et al., 1977). It also threatens biodiversity in invaded habitats in no fewer than 50 countries in temperate and tropical areas throughout the world, including countries in which it is a native species (Holm et al., 1977).</p> | remove India or give other countries and dates of introduction. | | |
| 15. | 9 | Technical | <p><i>Sorghum halepense</i> (Johnsongrass) is a perennial grass with a ribbed leaf sheath, conspicuous midrib, large, purplish panicles, and far-reaching rhizomes (Figures 1 and 2). It originated from the hybridization of <i>Sorghum arundinaceum</i> and <i>Sorghum propinquum</i> through chromosome doubling (chromosomes: $2n = 4x = 40$) (Ng'uni et al., 2010). <i>S. halepense</i> which is native to the Mediterranean area (Meredith, 1955) and was introduced to India in the late 1960s (Bor, 1960). It has become widespread, and is distributed from latitude 55° north to 45° south. It is best adapted to warm, humid areas with summer rainfall, areas with a high water table, and irrigated fields in subtropical zones. <i>S. halepense</i> is one of the most malignant weeds worldwide, impacting more than 30 cereal, vegetable and fruit crops (Holm et al., 1977). It also threatens</p> | It is a bit strange to focus on India here since the species has become established in all the warm regions of the world as explained in the next sentence. Either remove India or give other countries and dates of introduction. | European Union | <p>Incorporated (Modified).</p> <p>Text adjusted "other regions then" (see response above – comment 14)</p> |



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| | | | biodiversity in invaded habitats in no fewer than 50 countries in temperate and tropical areas throughout the world, including countries in which it is a native species (Holm <i>et al.</i> , 1977). | | | |
| 16. | 10 | Editorial | The main factors affecting the pest risk of <i>S. halepense</i> are that it: (1) has a high reproductive capacity; (2) is an alternate host of numerous pathogen species; (3) has allelopathic effects in and toxicity to livestock (da Nobrega <i>et al.</i>, 2006); (4) has developed resistance to a wide range of herbicide groups (Heap, n.d.); and (45) crosses with related species readily, which may produce more invasive hybrids and cause gene pollution of crop species (Arriola and Ellstrand, 1996). | Deletion of point (3): Is it normal/acceptable to make a direct reference to an animal health benefit in an ISPM? | EPPO | Considered, but not incorporated. This section is “Pest information” and <i>S. halepense</i> is known for its potential toxicity in livestock, which is related to food security, even though not directly related to IPPC mission (i.e. livestock). The pest risk, in general sense, is caused by its potential harmfulness. Here toxicity to livestock is one of the harmful factors of <i>S. halepense</i> |
| 17. | 10 | Editorial | The main factors affecting the pest risk of <i>S. halepense</i> are that it: (1) has a high reproductive capacity; (2) is an alternate host of numerous pathogen species; (3) has allelopathic effects in and toxicity to livestock (da Nobrega <i>et al.</i>, 2006); (4) has developed resistance to a wide range of herbicide groups (Heap, n.d.); and (45) crosses with related species readily, which may produce more invasive hybrids and cause gene pollution of crop species (Arriola and Ellstrand, 1996).species (Arriola and Ellstrand, 1996). | Deletion of point (3): Is it normal/acceptable to make a direct reference to an animal health benefit in an ISPM? | European Union | Considered, but not incorporated. See response above (comment 16). |
| 18. | 10 | Substantive | The main factors affecting the pest risk of <i>S. halepense</i> are that it: (1) has a high reproductive capacity <u>and the seeds have the characteristic of dormancy</u> ; (2) <u>The Sorghum halepense has strong competition ability and cause great yield lost of crop</u> ;(3) is an alternate host of numerous pathogen | The inference ability and seed dormancy of weed are important factors deciding its harmful level and environmental fitness. | China | Modified. |



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| | | | species; (34) has allelopathic effects in and toxicity to livestock (da Nobrega <i>et al.</i> , 2006); (45) has developed resistance to a wide range of herbicide groups (Heap, n.d.); and (56) crosses with related species readily, which may produce more invasive hybrids and cause gene pollution of crop species (Arriola and Ellstrand, 1996). | | | |
| 19. | 10 | Substantive | The main factors affecting the pest risk of <i>S. halepense</i> are that it: (1) has a high reproductive capacity; (2) is an alternate host of numerous pathogen species; (3) has allelopathic effects in and toxicity to livestock (da Nobrega <i>et al.</i> , 2006); (4) has developed resistance to a wide range of herbicide groups (Heap, n.d.); and (5) crosses with related species readily, which may produce more invasive hybrids and cause gene pollution of crop species (Arriola and Ellstrand, 1996). | Suggest substituting the term "gene pollution" with "gene introgression" (stable transfer of genetic material from one species/variety/population to another). This process is well known within the genus <i>Sorghum</i> (commercial sorghum, Johnson grass, shatter cane, and others). Indeed, genetic material is transferred between commercial sorghum and Johnson grass (in both directions). | United States of America | Incorporated. |
| 20. | 10 | Technical | The main factors affecting the pest risk of <i>S. halepense</i> are that it: (1) has a high reproductive capacity; (2) is an alternate host of numerous pathogen species; (3) has allelopathic effects in and toxicity to livestock (da Nobrega <i>et al.</i> , 2006); (4) has developed resistance to a wide range of herbicide groups (Heap, n.d.); and (5) readily crosses with related species (including crop species) readily, which may result in produce more invasive hybrids or and cause gene pollution of crop species (Arriola and Ellstrand, 1996). | Improvements to the English and highlighting that direct hybridisation with crop species is a risk. | EPPO, European Union | Modified. Text revised, but it doesn't need to extend of fix the range of related species because it is mentioned in the later sentence, |
| 21. | 10 | Technical | The main factors affecting the pest risk of <i>S. halepense</i> are that it: (1) has a high reproductive capacity; (2) is an alternate host of numerous pathogen species; (3) has allelopathic effects in and toxicity to livestock (da Nobrega <i>et al.</i> , 2006); (4) has developed resistance to a wide range of herbicide groups (Heap, n.d.); and (5) crosses with related species readily, which may produce more invasive hybrids and cause gene pollution of crop species (Arriola and Ellstrand, 1996). | Toxicity to livestock is not a factor affecting the pest risk. According to section 2.3.1. of ISPM 11 consequences considered should result from effects on plants. | COSAVE, Argentina, Peru, Brazil, Uruguay, | Considered but not incorporated. (See response to comment 16). |



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| 22. | 11 | Editorial | S. halepense is able to reproduce by rhizomes or seeds. Fragments of its long, vigorous and highly adaptable <u>Rhizomes</u> rhizome system readily sprout and can be distributed by tillage. An individual S. halepense plant is able to produce as many as 28 000 seeds in a growing season. These seeds are able to survive and germinate under most environmental conditions. Seed reproduction may generate diverse ecotypes that are distinct in morphology, anatomy and physiology. | 1) simplification of the English 2) Unnecessary wording. This is fairly basic biology and is applicable to any seed so does it really warrant specific mention? | EPPPO | Modified. |
| 23. | 11 | Editorial | <i>S. halepense</i> is able to reproduce by rhizomes or seeds. Fragments of its long, vigorous and highly adaptable <u>Rhizomes</u> rhizome system readily sprout and can be distributed by tillage. An individual <i>S. halepense</i> plant is able to produce as many as 28 000 seeds in a growing season. These seeds are able to survive and germinate under most environmental conditions. Seed reproduction may generate diverse ecotypes that are distinct in morphology, anatomy and physiology. | 1) Simplification of the English 2) Unnecessary wording. This is fairly basic biology and is applicable to any seed so does it really warrant specific mention? | European Union | Modified. |
| 24. | 12 | Editorial | Seeds are the main means of spread of <i>S. halepense</i> , and they are readily distributed by wind and water as well as by birds and other animals. More importantly, the seeds are frequently disseminated as a contaminant of commodities traded around the world; in particular, crop seeds and raw grains, such as <i>Sorghum bicolor</i> (sorghum), <i>Glycine max</i> (soybean), <i>Zea mays</i> (maize), <i>Triticum aestivum</i> (wheat) and <i>Sesamum indicum</i> (sesame), as well as forage, <i>Gossypium</i> spp. (cotton) and birdseed mixes. Therefore, seed quarantine is <u>key</u> the core task for the control of <i>S. halepense</i> , <u>and</u> which requires the prerequisite of accurate detection and identification <u>of seeds</u> . | Improved clarity. | EPPPO, European Union | Modified. |
| 25. | 12 | Technical | Seeds are the main means of spread of <i>S. halepense</i>, and they are readily distributed by wind and water as | This paragraph is not related to pest identification and additionally may lead to | COSAVE, | Modified. |



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| | | | well as by birds and other animals. More importantly, the seeds are frequently disseminated as a contaminant of commodities traded around the world; in particular, crop seeds and raw grains, such as <i>Sorghum bicolor</i> (sorghum), <i>Glycine max</i> (soybean), <i>Zea mays</i> (maize), <i>Triticum aestivum</i> (wheat) and <i>Sesamum indicum</i> (sesame), as well as forage, <i>Gossypium</i> spp. (cotton) and birdseed mixes. Therefore, seed quarantine is the core task for the control of <i>S. halepense</i>, which requires the prerequisite of accurate detection and identification. | the idea that it will be always necessary to establish measures for this pest even without an appropriate technical justification. | Argentina, Peru, Brazil, Uruguay, Chile, Paraguay | This paragraph was kept with modification to the last sentence. This paragraph is the prerequisite for developing this diagnostic protocol. The last sentence was removed as it was too directive to NPPOs. |
| 26. | 15 | Editorial | Synonyms: <i>Holcus halepensis</i> L., 1753 | Put the synonyms in alphabetical order unless there is a specific reason why they are not already (e.g. by how commonly they are used.) | EPPO, European Union | Modified |
| 27. | 16 | Technical | <i>Sorghum miliaceum</i> (Roxb.) Snowden, 1955 | <i>Sorghum miliaceum</i> is not a synonym of <i>Sorghum halepense</i> . In fact, they are different species. | Thailand | Incorporated |
| 28. | 17 | Technical | <i>Andropogon miliaceus</i> Roxb., 1820 | <i>Andropogon miliaceus</i> is not a synonym of <i>Sorghum halepense</i> . In fact, they are different species. <i>Andropogon miliaceus</i> is a synonym of <i>Sorghum miliaceum</i> . | Thailand | Incorporated |
| 29. | 18 | Technical | <i>Sorghum controversum</i> (Steud.) Snowden, 1955 | <i>Sorghum controversum</i> is not a synonym of <i>Sorghum halepense</i> . In fact, they are different species. | Thailand | Incorporated |
| 30. | 19 | Technical | <i>Andropogon controversus</i> Steud., 1854 | <i>Andropogon controversus</i> is not a synonym of <i>Sorghum halepense</i> . In fact, they are different species. <i>Andropogon controversus</i> is a synonym of <i>Sorghum controversum</i> . | Thailand | Incorporated |
| 31. | 29 | Editorial | Identification of <i>S. halepense</i> is commonly based on morphology. For suspected seeds with intact glumes and upper lemmas, morphological identification methods (section 4.1) are reliable. However, the fruits and seeds collected may be incomplete and | Editorial correction. | COSAVE, Argentina, Peru, Brazil, Uruguay, | Modified. |



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| | | | parts of their characters unclear. In such cases, molecular (section 4.2) or biochemical (section 4.3) identification methods may need to be used. Seeds may also be sown and grown into seedlings and then mature plants that can be morphologically (section 4.4) or cytologically (section 4.5) examined for taxonomic traits and subsequently identified. Figure 4 presents a flow diagram for the identification of <i>S. halepense</i> . | | | | Chile, Paraguay | | |
| 32. | 30 | Editorial | <i>S. halepense</i> is prone to be confused with five related species in the genus <i>Sorghum</i> : | | | | Put the five species into alphabetical order unless there is a specific reason why they are not already. | EPPO, European Union | Incorporated |
| 33. | 41 | Editorial | Species | Sessile spikelet | Caryopsis | Weight of 1 000 seeds (g, approximate) | According to the Barkworth, M.E. (2013), sessile spikelet bisexual is 3.8-6.5 mm long, 1.5-2.3 mm wide. | Japan | Incorporated |
| | | | <i>S. halepense</i> | Oval, (3.8) 4–5 (6.5-6) mm in length, appressed pubescent | Dark brown, obovate, 2.6–3.2 mm in length and 1.5–1.8 mm in width | 4.9 | | | |
| | | | <i>S. xalimum</i> | Oval to oblong, 4.5–6 mm in length, short pubescent | Red–brown, broadly ovate or oval, 3.3–4 mm in length and 2–2.3 mm in width | 6.6 | | | |
| | | | <i>S. propinquum</i> | Oval to oblong, 3.8–4.5 mm in length, bearded | Brown, broadly ovate or broadly oval, approximately 2 mm in length and 1.5 mm in width | 3.8 | | | |
| | | | <i>S. sudanense</i> | Oval, (5) 6–8 mm in length, sparsely pubescent | Red–brown, broadly ovate, 3.5–4.5 mm in length, 2.5–2.8 mm in width | 10–15 | | | |



| | | | <table border="1"> <tr> <td><i>S. bicolor</i> or</td> <td>Elliptic to oblong or ovate, (3) 4.5– 6 (9) mm in length, densely hispid, or pubescent to glabrous</td> <td>Pink to red–brown, ovate, 3.5–4 mm in length, 2.5–3 mm in width</td> <td>>20</td> </tr> <tr> <td><i>Sorghum</i> spp. hybrid cv. Silk</td> <td>Oval, approximately 3.8 mm in length, short pubescent</td> <td>Yellow or yellow–brown, broadly ovate, 2.5–4 mm in length and 1.7–2.5 mm in width</td> <td>4.2</td> </tr> </table> | <i>S. bicolor</i> or | Elliptic to oblong or ovate, (3) 4.5– 6 (9) mm in length, densely hispid, or pubescent to glabrous | Pink to red–brown, ovate, 3.5–4 mm in length, 2.5–3 mm in width | >20 | <i>Sorghum</i> spp. hybrid cv. Silk | Oval, approximately 3.8 mm in length, short pubescent | Yellow or yellow–brown, broadly ovate, 2.5–4 mm in length and 1.7–2.5 mm in width | 4.2 | | | | | | | |
|-------------------------------------|--|---|---|----------------------|--|---|--|-------------------------------------|--|---|---|--|--|-----------------------------|--|--|-------|---|
| <i>S. bicolor</i> or | Elliptic to oblong or ovate, (3) 4.5– 6 (9) mm in length, densely hispid, or pubescent to glabrous | Pink to red–brown, ovate, 3.5–4 mm in length, 2.5–3 mm in width | >20 | | | | | | | | | | | | | | | |
| <i>Sorghum</i> spp. hybrid cv. Silk | Oval, approximately 3.8 mm in length, short pubescent | Yellow or yellow–brown, broadly ovate, 2.5–4 mm in length and 1.7–2.5 mm in width | 4.2 | | | | | | | | | | | | | | | |
| 34. | 41 | Technical | <table border="1"> <thead> <tr> <th>Species</th> <th>Sessile spikelet</th> <th>Caryopsis</th> <th>Weight of 1 000 seeds (g, approximate)</th> </tr> </thead> <tbody> <tr> <td><i>S. halepense</i></td> <td>Oval <u>elliptic or ovate</u>, (3.8) 4–5 (5.6) mm in length, appressed pubescent</td> <td>Dark brown, obovate to elliptic,</td> <td>2.6–3.2 mm in length and 1.5–1.8 mm in width 4.9</td> </tr> <tr> <td></td> <td><u>elliptic to oblong</u> Oval to oblong, 4.5–</td> <td>Red–brown, broadly ovate or</td> <td></td> </tr> </tbody> </table> | Species | Sessile spikelet | Caryopsis | Weight of 1 000 seeds (g, approximate) | <i>S. halepense</i> | Oval <u>elliptic or ovate</u> , (3.8) 4–5 (5.6) mm in length, appressed pubescent | Dark brown, obovate to elliptic , | 2.6–3.2 mm in length and 1.5–1.8 mm in width 4.9 | | <u>elliptic to oblong</u> Oval to oblong , 4.5– | Red–brown, broadly ovate or | | <p>Oval means the width over one-half of the length. Based on the samples and pictures of <i>S. halepense</i>, also based on some relative references (Flora of China Editorial Committee. 2013. Poaceae Flora of China, 22 URL: http://foc.eflora.cn/content.aspx?TaxonId=130722), the sessile spikelet of <i>S. halepense</i> is not oval in most cases. For the same reasons, some morphological descriptions of other species ere been suggested to confirm and revise.</p> | China | Incorporated. In addition, revised description on sessile spikelet because based on “Flora of China” it is elliptic |
| Species | Sessile spikelet | Caryopsis | Weight of 1 000 seeds (g, approximate) | | | | | | | | | | | | | | | |
| <i>S. halepense</i> | Oval <u>elliptic or ovate</u> , (3.8) 4–5 (5.6) mm in length, appressed pubescent | Dark brown, obovate to elliptic , | 2.6–3.2 mm in length and 1.5–1.8 mm in width 4.9 | | | | | | | | | | | | | | | |
| | <u>elliptic to oblong</u> Oval to oblong , 4.5– | Red–brown, broadly ovate or | | | | | | | | | | | | | | | | |



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|--|--|--|-------------------------------------|---|--|-------|--|--|--|
| | | | <i>S. xal- mum</i> | 6 mm in length, short pubescent | oval, 3.3– 4 mm in length and 2–2.3 mm in width | 6.6 | | | |
| | | | <i>S. propinquum</i> | 4.5 mm in length, bearded | elliptic to oblong to oblong , 3.8– Brown, broadly ovate or broadly oval, approximately 2 mm in length and 1.5 mm in width | 3.8 | | | |
| | | | <i>S. sudanense</i> | Oval, (5) 6–8 mm in length, sparsely pubescent | Red–brown, broadly ovate, 3.5–4.5 mm in length, 2.5–2.8 mm in width | 10–15 | | | |
| | | | <i>S. bicolor</i> | Elliptic to oblong or ovate, (3) 4.5–6 (9) mm in length, densely hispid, or pubescent to glabrous | Pink to red–brown, ovate, 3.5–4 mm in length, 2.5–3 mm in width | >20 | | | |
| | | | <i>Sorghum</i> spp. hybrid cv. Silk | Oval, approximately 3.8 mm in | Yellow or yellow–brown, broadly ovate, 2.5–4 mm in length and 1.7– | 4.2 | | | |



| | | length, short pubescent | | 2.5 mm in width | | | | | |
|-----|----|-------------------------|----------------------|---|--|--|---|----------------------|--------------|
| 35. | 41 | Technical | Species | Sessile spikelet | Caryopsis | Weight of 1 000 seeds (g, approximate) | For sessile spikelet, it would be useful to provide length and width. According to Flora of North America, the length of sessile spikelet for <i>S. halapense</i> can reach 6.5 mm. For <i>S. x almum</i> , <i>S. propinquum</i> and <i>S. bicolor</i> , Clayton et al. (2006) provide higher value. Clayton, W.D., Vorontsova, M.S., Harman, K.T. and Williamson, H. (2006 onwards). GrassBase - The Online World Grass Flora. http://www.kew.org/data/grasses-db.html . | EPPO, European Union | Incorporated |
| | | | <i>S. halepense</i> | Oval, (3.8) 4–5 (6.5 5.6) mm in length, appressed pubescent | Dark brown, obovate, 2.6–3.2 mm in length and 1.5–1.8 mm in width | 4.9 | | | |
| | | | <i>S. x almum</i> | Oval to oblong, 4.5–6.5 mm in length, short pubescent | Red–brown, broadly ovate or oval, 3.3– 4 mm in length and 2–2.3 mm in width | 6.6 | | | |
| | | | <i>S. propinquum</i> | Oval to oblong, 3.8– 5.4.5 mm in length, bearded | Brown, broadly ovate or broadly oval, approximately 2 mm in length and 1.5 mm in width | 3.8 | | | |
| | | | <i>S. sudanense</i> | Oval, (5) 6–8 mm in length, sparsely pubescent | Red–brown, broadly ovate, 3.5–4.5 mm in length, 2.5–2.8 mm in width | 10–15 | | | |



| | | | <table border="1"> <tr> <td><i>S. bicolor</i></td> <td>Elliptic to oblong or ovate (3) 4.5–6 (109) mm in length, sparsely pubescent, densely hispid, or pubescent to glabrous</td> <td>Pink to red–brown, ovate, 3.5–4 mm in length, 2.5–3 mm in width</td> <td>>20</td> </tr> <tr> <td><i>Sorghum</i> spp. hybrid cv. Silk</td> <td>Oval, approximately 3.8 mm in length, short pubescent</td> <td>Yellow or yellow–brown, broadly ovate, 2.5–4 mm in length and 1.7–2.5 mm in width</td> <td>4.2</td> </tr> </table> | <i>S. bicolor</i> | Elliptic to oblong or ovate (3) 4.5–6 (109) mm in length, sparsely pubescent, densely hispid, or pubescent to glabrous | Pink to red–brown, ovate, 3.5–4 mm in length, 2.5–3 mm in width | >20 | <i>Sorghum</i> spp. hybrid cv. Silk | Oval, approximately 3.8 mm in length, short pubescent | Yellow or yellow–brown, broadly ovate, 2.5–4 mm in length and 1.7–2.5 mm in width | 4.2 | | | | | | | | |
|-------------------------------------|--|---|---|-------------------|--|---|-------------|-------------------------------------|---|---|----------|-------------------|--|--|----------|---|---|-------|--------------|
| <i>S. bicolor</i> | Elliptic to oblong or ovate (3) 4.5–6 (109) mm in length, sparsely pubescent, densely hispid, or pubescent to glabrous | Pink to red–brown, ovate, 3.5–4 mm in length, 2.5–3 mm in width | >20 | | | | | | | | | | | | | | | | |
| <i>Sorghum</i> spp. hybrid cv. Silk | Oval, approximately 3.8 mm in length, short pubescent | Yellow or yellow–brown, broadly ovate, 2.5–4 mm in length and 1.7–2.5 mm in width | 4.2 | | | | | | | | | | | | | | | | |
| 36. | 44 | Technical | <table border="1"> <thead> <tr> <th>Glume</th> <th>Lower glume</th> <th>Upper glume</th> <th>Upper lemma</th> </tr> </thead> <tbody> <tr> <td><i>S. halepense</i></td> <td>Subleathery, tawny, red–brown, or purple–black</td> <td>Apex clearly tridentical at e, 5–7-veined, dorsum ciliary but the rest glabrous</td> <td>3-veined</td> </tr> <tr> <td><i>S. xalimum</i></td> <td>Chartaceous or subleathery, dark brown</td> <td>Apex little tridentical at e, 5–7-veined, dorsum ciliary but the rest glabrous</td> <td>3-veined</td> </tr> </tbody> </table> | Glume | Lower glume | Upper glume | Upper lemma | <i>S. halepense</i> | Subleathery, tawny, red–brown, or purple–black | Apex clearly tridentical at e, 5–7-veined, dorsum ciliary but the rest glabrous | 3-veined | <i>S. xalimum</i> | Chartaceous or subleathery, dark brown | Apex little tridentical at e, 5–7-veined, dorsum ciliary but the rest glabrous | 3-veined | <p>Triangular lanceolate, apex bilobed and awned or not; awn 10–16 mm</p> <p>Lanceolate, apex obtuse or slightly acute, bilobed, awned; awn approximately 15 mm</p> | <p>The glume of <i>S. halepense</i> is subleathery. And the upper lemma of <i>S. sudanense</i> is ovate or elliptic. Based on some relative references (Flora of China Editorial Committee. 2013. Poaceae Flora of China, 22 URL: http://foc.eflora.cn/content.aspx?TaxonId=130722), some morphological descriptions of the two species are suggested to confirm and revise.</p> | China | Incorporated |
| Glume | Lower glume | Upper glume | Upper lemma | | | | | | | | | | | | | | | | |
| <i>S. halepense</i> | Subleathery, tawny, red–brown, or purple–black | Apex clearly tridentical at e, 5–7-veined, dorsum ciliary but the rest glabrous | 3-veined | | | | | | | | | | | | | | | | |
| <i>S. xalimum</i> | Chartaceous or subleathery, dark brown | Apex little tridentical at e, 5–7-veined, dorsum ciliary but the rest glabrous | 3-veined | | | | | | | | | | | | | | | | |



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|-------------------------------------|---|--|---|--|---|---|----------|--|---------------------|-------------------------------------|---|-----------------------------|--|-------------------|-----------------------------|--|------------|--|-------------------------------------|--|--|----------|--|--|--|--|
| | | | <table border="1"> <tr> <td><i>S. propinquum</i></td> <td>Subleathery, dark brown with inconspicuous crossveins</td> <td>9–11-veined, apex acute to apiculate or tridenticulate, pubescent</td> <td>7-veined</td> <td>Lanceolate, approximately 3.5 mm in length, acute or emarginate, awnless</td> </tr> <tr> <td><i>S. sudanense</i></td> <td>Leathery, lemon yellow to red–brown</td> <td>Apex bidenticulate, 11–13-veined, usually with crossveins, dorsum short ciliary</td> <td>5–7-veined, with crossveins</td> <td>ovate or elliptic Ovate or oval, apex bilobed, awned; awn 10–16 mm</td> </tr> <tr> <td><i>S. bicolor</i></td> <td>Leathery, pink to red–brown</td> <td>Apex acute or tridenticulate, 12–16-veined with crossveins, dorsum dense ciliary</td> <td>7–9-veined</td> <td>Lanceolate to long oval, 2–4-veined, apex bilobed, awned; awn approximately 1 mm</td> </tr> <tr> <td><i>Sorghum</i> spp. hybrid cv. Silk</td> <td>Leathery, tawny, red–brown or purple–black</td> <td>Apex little tridenticulate, 5–7-veined, dorsum ciliary but</td> <td>3-veined</td> <td>Broad lanceolate, apex slightly bilobed, awnless</td> </tr> </table> | <i>S. propinquum</i> | Subleathery, dark brown with inconspicuous crossveins | 9–11-veined, apex acute to apiculate or tridenticulate, pubescent | 7-veined | Lanceolate, approximately 3.5 mm in length, acute or emarginate, awnless | <i>S. sudanense</i> | Leathery, lemon yellow to red–brown | Apex bidenticulate, 11–13-veined, usually with crossveins, dorsum short ciliary | 5–7-veined, with crossveins | ovate or elliptic Ovate or oval , apex bilobed, awned; awn 10–16 mm | <i>S. bicolor</i> | Leathery, pink to red–brown | Apex acute or tridenticulate, 12–16-veined with crossveins, dorsum dense ciliary | 7–9-veined | Lanceolate to long oval, 2–4-veined, apex bilobed, awned; awn approximately 1 mm | <i>Sorghum</i> spp. hybrid cv. Silk | Leathery, tawny, red–brown or purple–black | Apex little tridenticulate, 5–7-veined, dorsum ciliary but | 3-veined | Broad lanceolate, apex slightly bilobed, awnless | | | |
| <i>S. propinquum</i> | Subleathery, dark brown with inconspicuous crossveins | 9–11-veined, apex acute to apiculate or tridenticulate, pubescent | 7-veined | Lanceolate, approximately 3.5 mm in length, acute or emarginate, awnless | | | | | | | | | | | | | | | | | | | | | | |
| <i>S. sudanense</i> | Leathery, lemon yellow to red–brown | Apex bidenticulate, 11–13-veined, usually with crossveins, dorsum short ciliary | 5–7-veined, with crossveins | ovate or elliptic Ovate or oval , apex bilobed, awned; awn 10–16 mm | | | | | | | | | | | | | | | | | | | | | | |
| <i>S. bicolor</i> | Leathery, pink to red–brown | Apex acute or tridenticulate, 12–16-veined with crossveins, dorsum dense ciliary | 7–9-veined | Lanceolate to long oval, 2–4-veined, apex bilobed, awned; awn approximately 1 mm | | | | | | | | | | | | | | | | | | | | | | |
| <i>Sorghum</i> spp. hybrid cv. Silk | Leathery, tawny, red–brown or purple–black | Apex little tridenticulate, 5–7-veined, dorsum ciliary but | 3-veined | Broad lanceolate, apex slightly bilobed, awnless | | | | | | | | | | | | | | | | | | | | | | |
| 37. | 60 | Editorial | In this diagnostic protocol, methods (including | Delete unnecessary brackets | Canada | Incorporated/Modified. | | | | | | | | | | | | | | | | | | | | |



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|-----|----|-----------|--|--|---|--|
| | | | reference to brand names) are described as published, as these defined the original level of sensitivity, specificity and/or reproducibility achieved. The use of names of reagents, chemicals or equipment in these diagnostic protocols implies no approval of them to the exclusion of others that may also be suitable. (This information is given for the convenience of users of this protocol and does not constitute an endorsement by the CPM of the chemical, reagent and/or equipment named). Laboratory procedures presented in the protocols may be adjusted to the standards of individual laboratories, provided that they are adequately validated. | | | Disclaimer adjusted to the standard text in all diagnostic protocols. |
| 38. | 60 | Technical | In this diagnostic protocol, methods (including reference to brand names) are described as published, as these defined the original level of sensitivity, specificity and/or reproducibility achieved. The use of names of reagents, chemicals or equipment in these diagnostic protocols implies no approval of them to the exclusion of others that may also be suitable. (This information is given for the convenience of users of this protocol and does not constitute an endorsement by the CPM of the chemical, reagent and/or equipment named). Laboratory procedures presented in the protocols may be adjusted to the standards of individual laboratories, provided that they are adequately validated. <u>Under certain circumstances, seedling from seed samples may also be used to extract DNA</u> | If there is only a small number of seeds and they are vigorous, the quality of DNA extracted from seedlings is relatively higher than only from seeds. | China | Modified. According to instructions to authors and agreement by the SC (and the according to the recent adopted DPs), this disclaimer is in the main text and in footnotes (if a brand name in the protocol). However, the suggestion was incorporated in paragraph 62 (section “4.2.1 Methods based on DNA markers”) |
| 39. | 60 | Technical | In this diagnostic protocol, methods (including reference to brand names) are described as published, as these defined the original level of sensitivity, specificity and/or reproducibility achieved. The use of names of reagents, chemicals or equipment in these diagnostic protocols implies no approval of them to the exclusion of others that may also be suitable. (This information is given for the convenience of users of this protocol and does not constitute an endorsement by the CPM of the | Texted deleted and included in the footnote as previously agreed. | COSAVE, Argentina, Peru, Brazil, Uruguay, Chile, Paraguay | Modified. According to instructions to authors and agreement by the SC (and according to the recent adopted DPs), this disclaimer is in the main text and in footnotes (if a brand name in the protocol). |



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| | | | <p>chemical, reagent and/or equipment named.</p> <p>Laboratory procedures presented in the protocols may be adjusted to the standards of individual laboratories, provided that they are adequately validated.</p> | | | |
| 40. | 62 | Substantive | <p>For DNA extraction from seed samples, refer to the source paper of the molecular method for the specific technique used (Chen et al., 2009). The method described by Moller <i>et al.</i> (1992) is recommended for DNA microextraction from seeds of <i>Sorghum</i> species. Laboratories may find that alternative DNA extraction techniques work equally well. If more than one seed is included in the extraction, the DNA may comprise a mixture of species.</p> <p>Note: it would be better if there are recommended protocol for DNA extraction for this guidelines</p> | if it is possible, it would be better if there are included recommended method for DNA extraction, for guidelines to conduct appropriate DNA extraction especially for this species | Indonesia | <p>Modified.</p> <p>The DP refers to literature DNA extraction methods. A footnote was included to mention that Laboratories may find that alternative DNA extraction techniques work equally well</p> |
| 41. | 106 | Technical | 4.4 Morphological identification of plants | It would be useful to have some estimated timeframes | Australia | <p>Incorporated.</p> <p>Added "for more than 100 days" in which seeds can be grown.</p> |
| 42. | 110 | Technical | <p>Mature plant: Perennial with vigorous, spreading rhizomes. Culms 0.5–1.5 (–32.0) m tall, 4–6 (–20) mm in diameter; nodes puberulous. Leaf sheaths glabrous; leaf blades linear or linear-lanceolate, (10–) 25–80 (–</p> <p>90) × (0.58–) 1–4 cm, glabrous; ligule 0.5–1 (2–6) mm, glabrous ciliate <u>membrane</u>.</p> | Change of mimum or maximum size according to Clayton et al. (2006). | EPPO | Incorporated. |
| 43. | 110 | Technical | <p>Mature plant: Perennial with vigorous, spreading rhizomes. Culms 0.5–1.5 (–32.0) m tall, 4–6 (–20) mm in diameter; nodes puberulous. Leaf sheaths glabrous; leaf blades linear or linear-lanceolate,</p> | Change of mimum or maximum size according to Clayton et al. (2006). | European Union | Incorporated. |



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| | | | (10–) 25–80 (– 90) × (0.58–) 1–4 cm, glabrous; ligule 0.5–1 (2–6) mm, glabrous ciliolate membrane. | | | |
| 44. | 111 | Technical | Inflorescence: Panicle lanceolate to pyramidal in outline, (10–) 20–40 (–559) cm, soft white hairs in basal axil; primary branches solitary or whorled, spreading, lower part bare, upper part branched, the secondary branches tipped by racemes; racemes fragile, composed of (1–) 2–5 spikelet pairs. | Maximum size changed according to Clayton et al (2006) | EPPO, European Union | Incorporated. |
| 45. | 112 | Technical | Spikelet: Usually in pairs although towards the tip of the inflorescence they may occur in threes; when the spikelet is in pairs, the lower is sessile and perfect with the upper, pedicelled, narrow, long and stamen-bearing; when the spikelet is in threes, one is sessile and perfect, the others are pedicelled and staminate. Sessile spikelet elliptic, (3.8–) 4–5 (–6.5) mm; callus obtuse, bearded; subleathery lower glume leathery, often pale yellow or yellowish brown at maturity, shortly pubescent or glabrescent, 5–7-veined, veins distinct in upper part, apex tridenticulate; upper lemma acute and mucronate or bilobed and awned or not; awn 1–1.6 cm. Pedicelled spikelet staminate, narrowly lanceolate, (3.6–) 4.5–7 mm, often violet-purple. | The glume of <i>S. halepense</i> is subleathery. | China | Incorporated. |
| 46. | 119 | Substantive | – Culm base 3–9 mm in diameter <i>S. sudanense</i> | <i>Sorghum bicolor</i> subsp. <i>arundinaceum</i> (Desv.) de Wet & J.R. Harlan should be included in the key. | EPPO, European Union | Considered, but not incorporated. <i>Sorghum bicolor</i> subsp. <i>arundinaceum</i> (Desv.) de Wet & J.R. Harlan is a wild species in <i>Sorghum</i> and has a limited distribution range. So it may not be often found in traded commodities The key is for the five main |



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| | | | | | | species that are referred to elsewhere in the DP. (i.e. it does not cover all potentially confusing species.) |
| 47. | 145 | Technical | <p>The mean fluorescence of nuclei is quantified using a flow cytometer (Coulter Electronics)¹ equipped with a water- cooled laser tuned at 514 nm and 500 mW. Fluorescence at >615 nm is detected with a photomultiplier screened by a long pass filter. The mean 2C DNA content of each target species is calculated by comparing its mean nuclear fluorescence with the mean nuclear fluorescence of an internal standard. Because of the variation of <i>Sorghum</i> DNA content, one of two different internal standards is used to avoid overlap of the standard and target species. One standard, <i>Arabidopsis thaliana</i> ecotype Columbia, has a genome size of 157 Mb or</p> <p>1C = 0.16 pg. The DNA content of <i>A. thaliana</i> and <i>S. bicolor</i> Tx623 (2C DNA content = 1.67 pg) is determined from 15 replicates of leaf samples from <i>S. bicolor</i> and <i>A. thaliana</i> Columbia. At least three replicates for each test sample are analysed to obtain the mean DNA content (Price <i>et al.</i>, 2005; Jessup <i>et al.</i>, 2012).</p> | <p>The following footnote should be inserted: “The use of brand names of reagents, chemicals or equipment in this diagnostic protocol implies no approval of them to the exclusion of others that may also be suitable. This information is given for the convenience of users of this protocol and does not constitute an endorsement by the CPM of the chemical, reagent and/or equipment named. Equivalent products may be used if they can be shown to lead to the same results.</p> | <p>COSAVE, Argentina, Peru, Brazil, Uruguay, Chile, Paraguay</p> | Incorporated. |
| 48. | 155 | Editorial | <p>A request for a revision to a diagnostic protocol may be submitted by national plant protection organizations (NPPOs), regional plant protection organizations (RPPOs) or Commission on Phytosanitary Measures (CPM) subsidiary bodies through the IPPC Secretariat (ippc@fao.org), which will in turn forward it to the Technical Panel on Diagnostic Protocols (TPDP).</p> | <p>Add "A" at the beginning of the sentence.</p> | <p>Canada</p> | Incorporated. |
| 49. | 164 | Editorial | <p>CSIRO. 1978. <i>Sorghum</i> spp. hybrid (forage</p> | <p>Add a reference : Clayton, W.D.,</p> | <p>EPPO</p> | Incorporated |



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| | | | sorghum hybrids) cv. Silk. <i>Journal of the Australian Institute of Agricultural Science</i> , 44(3 and 4): 219–221. | Vorontsova, M.S., Harman, K.T. and Williamson, H. (2006 onwards). GrassBase - The Online World Grass Flora. http://www.kew.org/data/grasses-db.html . | | |
| 50. | 164 | Editorial | CSIRO . 1978. <i>Sorghum</i> spp. hybrid (forage sorghum hybrids) cv. Silk. <i>Journal of the Australian Institute of Agricultural Science</i> , 44(3 and 4): 219–221. | Add a reference: Clayton, W.D., Vorontsova, M.S., Harman, K.T. and Williamson, H. (2006 onwards). GrassBase - The Online World Grass Flora. http://www.kew.org/data/grasses-db.html . | European Union | Incorporated |